CRISPR/Cas9 Targeted Disruption of Herpes Simplex Virus type 1 in a Rabbit Latency Model Reduces Viral Reactivation and Associated Corneal Pathology

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Introduction

• Recurrent Herpes Simplex Virus-1 (HSV-1) stromal keratitis is considered the leading cause of infectious corneal blindness worldwide¹,², despite the current standard of care³.
• Recurrent HSV-1 ocular keratitis results from reactivation of latent virus within the trigeminal ganglia (TG), followed by an inflammatory response to viral infection within the stroma¹.
• To address this unmet medical need, we have developed a gene editing approach using Adeno-Associated Virus (AAV)-delivered CRISPR-associated protein-9 (Cas9) nucleases from S. aureus (SaCas9) and guide RNAs (gRNAs) to target latent HSV-1 genomes.
• The objective of this study was to evaluate the efficacy of the CRISPR/Cas9 gene editing system in a rabbit model of recurrent HSV-1 keratitis.

Methods

gRNA selection
• gRNAs (N=411) and SaCas9 assembled into ribonucleoproteins (RNPs) were screened against HSV-1-specific sequences in a high-throughput in vitro screen.
• Benchmark gRNAs were characterized in a viral replication assay. (Figure 1)

Characterization of benchmark gRNAs
• Benchmark gRNAs were characterized in a viral replication assay using RNPs complexed with individual gRNAs. Briefly, Vero cells were transfected with RNPs, and 24 hrs later, challenged with HSV-1 175syn across a range of Multiplicities-of-Infection (MOIs). At 24 hrs post infection, levels of viral genomes in supernatants were quantified by qPCR. Percent inhibition of viral replication was calculated based on the AUC of the MOI dose response curves, relative to that of a non-targeting control gRNA. (Figure 2)

Rabbit model of HSV-1 keratitis
• HSV-1 175syn was applied topically to scared corneas of New Zealand White rabbits to establish latency in the TG in 3.5 weeks. AAV expressing SaCas9 and gRNAs were dosed via corneal abrasion and allowed to express the transgenes for 4 weeks prior to HSV-1 reactivation by epinephrine iontophoresis on rabbit corneas.
• Slit lamp examinations (SLE) were conducted every other day to monitor symptoms of HSV-1 reactivation. HSV-1 viral production was determined by a plaque assay of daily tear samples (Figure 4).
• TG were harvested 10 days post induction. HSV-1 and AAV viral genomes in TG were quantified by qPCR. Statistical analyses utilized one-way ANOVA (Figures 7 & 8).

Study I: in vitro gRNA Selection

FIGURE 1. Design of the in vitro RNP Selection.

FIGURE 2. gRNAs targeting UL48 and RL2 showed the greatest efficacy in inhibiting viral replication.

Study II: in vivo Efficacy Model

Table 1. Treatment groups for in vivo testing.

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FIGURE 3. AAV vector map.

FIGURE 5. Corneal lesions suppressed by up to 91% with a dual-gRNA approach.

FIGURE 4. HSV-1 reactivation rabbit model study design.

FIGURE 6. Infectious HSV-1 in tear films inhibited up to 75% by a dual-gRNAs approach.

FIGURE 7. CRISPR/Cas9 or acyclovir had little impact on HSV-1 copy numbers in TG.

FIGURE 8. AAV copy numbers in TGs were not above background.

Conclusions

1. An in vitro gRNA selection method was developed to screen gRNAs that target essential HSV genes for viral replication.
2. In vitro characterization demonstrated gRNAs targeting UL48 and RL2 had the greatest efficiency in inhibiting viral replication.
3. Evaluation of gRNAs targeting UL48 and RL2 in the rabbit reactivation model indicated that the combination of gRNAs resulted in the greatest efficacy.
4. In summary, we have identified gRNAs and demonstrated their ability to reduce viral shedding and corneal lesion development in a rabbit model of HSV-1 keratitis and reactivation. The impact of the gRNAs on viral load and pathology may reflect activity at the corneal surface.

Reference: